

**Protocol for Nail Plate Fragments Histologic Processing and PAS Staining**

1. Place a small amount of gelatin in water bath.
2. Cut ribbon at 4 µm and float onto gelatin water bath.
3. Using positively charged slide, pick up desired sections for PAS and H&E slides.
4. Place in 65° C oven for 45 min (more time for “difficult” specimens).
5. Deparaffinize slides using xylene or xylene substitutes and hydrate through alcohols (or place PAS slides on programmed deparaffin run and H&E slides on H&E program on stainer).
6. Gently rinse slide in running tap water.
7. Rinse slide in distilled water.
8. Do not digest the slides.
9. Place slide in 1% periodic acid for 10 min.
10. Gently rinse slide quickly in distilled water.
11. Place slide in Schiff solution for 10 min.
12. Gently rinse slides in warm–hot tap water for 5 min.
13. Place slide in light green stain as needed to reach desired background intensity.  
(approximately 30 sec)
14. Dehydrate slide through three changes of absolute alcohol.
15. Clear slide through three changes of xylene or xylene substitute.
16. Coverslip using permanent mounting media.